

¹⁹F Magnetic Resonance Spectroscopic Investigation of the Binding of *N*-Trifluoroacetylated Amino-acids by Chymotrypsin

By H. ASHTON and B. CAPON*

(Chemistry Department, Glasgow University, Glasgow W.2)

Summary The binding of *N*-trifluoroacetyl-D-tryptophan by chymotrypsin causes a downfield shift in the ¹⁹F n.m.r. signal in the pH range 6.33—8.12 and that of *N*-trifluoroacetyl-D-phenylalanine and of *N*-trifluoroacetyl-L-tryptophan causes a downfield shift at pH 6.34.

THE shift in the ¹⁹F n.m.r. signal of the sodium salt of *N*-trifluoroacetyl-D-tryptophan in the presence of chymotrypsin may be used to measure the dissociation constant of the complex formed by this inhibitor and the enzyme (K_I). Some of the values obtained are given in the Table. At all pH values there was a variation in δ of more than 4 Hz over the concentration range 5×10^{-2} — 5×10^{-3} M with the concentration of chymotrypsin 2×10^{-3} M. At some pH values, *e.g.* 6.33, this variation was as large as 13.4 Hz. The direction of the shift was always downfield. The large increase in K_I at pH *ca.* 8 is similar to that reported by Johnson and Knowles¹ for the binding of *N*-acetyl-D-tryptophan to chymotrypsin, measured by equilibrium dialysis. Within experimental error (± 0.1 Hz) the chemical shift was independent of concentration in the

absence of enzyme. Care was taken to keep the pH constant over the whole concentration range studied

TABLE

Binding of N-trifluoroacetyl-D-tryptophan to chymotrypsin

pH	6.33	6.89	7.72	8.12
$10^3 K_I$ (M) ^a	0.65	1.59	2.86	7.2
Δ (Hz) ^b	112	182	148	255
K_I (M) ^c	<i>ca.</i> 10^{-3}		<i>ca.</i> 10^{-2}	

^a Dissociation constant of chymotrypsin-*N*-trifluoroacetyl-D-tryptophan complex determined from ¹⁹F-resonance results at 94.1 MHz; trifluoroacetic acid was used as external standard. ^b Difference in chemical shift between bound and unbound *N*-trifluoroacetyl-D-tryptophan. ^c Kinetically determined inhibition constant.

(always to within ± 0.05). The values of K_I in the Table were calculated by the method of Nakano, Nakano, and Higuchi² and are in moderate agreement with those determined kinetically for the hydrolysis of *N*-furylacryl-L-tryptophanamide.

There have been two previous investigations of the binding of *N*-trifluoroacetyl-amino-acids to chymotrypsin by ^{19}F resonance. Zeffren and Reavill reported that in citrate buffer (0.1M), at pH 6.0, there was a downfield shift of the ^{19}F signal of one enantiomer of *N*-trifluoroacetyl-DL-phenylalanine in the presence of chymotrypsin³ but Sykes reported an upfield shift in the ^{19}F signal of *N*-trifluoroacetyl-D-phenylalanine in 0.1M-tris-HCl buffer at pH 7.8.⁴ However the shifts observed were very small. Thus, the reported variation in shift for a concentration range $4 \times 10^{-3}\text{ M}$ — $3.6 \times 10^{-2}\text{ M}$ was less than 1 Hz with an experimental error of $\pm 0.1\text{ Hz}$.⁴ In our experiments the difference in pH between a solution of 4×10^{-3} and $3.6 \times 10^{-2}\text{ M}$ *N*-trifluoroacetyl-D-phenylalanine in the presence of chymotrypsin ($2 \times 10^{-3}\text{ M}$) in a 0.1 M-tris-HCl buffer of nominal pH 7.88 was over one pH unit. The difference between chemical shifts in the presence and absence of chymotrypsin was $4.5 \pm 0.2\text{ Hz}$ for all concentrations of *N*-trifluoroacetyl-D-phenylalanine and varied randomly. The signal moved to lower field in the presence of chymotrypsin. The ^{19}F signal of the sodium salt of *N*-trifluoroacetyl-D-phenylalanine at pH 7.96 ± 0.01 moved to lower field in the presence of chymotrypsin ($2 \times 10^{-3}\text{ M}$) but the

shift was constant at $3.2 \pm 0.2\text{ Hz}$ over the concentration range 5×10^{-3} — $4 \times 10^{-2}\text{ M}$. At pH 6.34 (± 0.05) there was a downfield shift in the presence of chymotrypsin which varied with concentration. The value of K_1 was calculated to be $4.3 \times 10^{-2}\text{ M}$, in quite good agreement with that determined kinetically (3×10^{-2}).

The ^{19}F signal of *N*-trifluoroacetyl-L-tryptophan at pH 6.34 shows a concentration-dependent downfield shift in the presence of chymotrypsin but this is too small ($< 4\text{ Hz}$) to allow a reliable value of K_1 to be determined.

It has been concluded⁵ from the reports that the ^{19}F signal of *N*-trifluoroacetyl-D-phenylalanine shows a downfield shift in the presence of chymotrypsin at pH 6.0³ and an upfield shift at pH 7.8⁴ that there is a 'major change in the magnetic environment of the fluorine atoms of the bound inhibitor with pH'. The results obtained in this investigation suggest that if there is such a change the effect on the ^{19}F -shift of the bound inhibitor is not so large as previously supposed.

We thank Mr. J. Gall for measurements of the ^{19}F chemical shifts.

(Received, February 22nd, 1971; Com. 150.)

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⁵ C. G. K. Roberts and O. Jardetzky, *Adv. Protein Chem.*, 1970, **24**, 512.